

Applications of Microfluidics in Waterborne Pathogen Monitoring

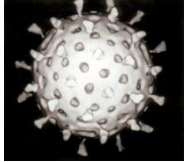

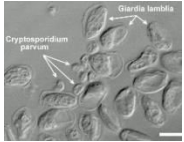
Dr Helen Bridle

Monitoring for Waterborne Pathogens

- Why?
- Health and economic impacts
- Outbreaks
- Endemic disease
- Food production

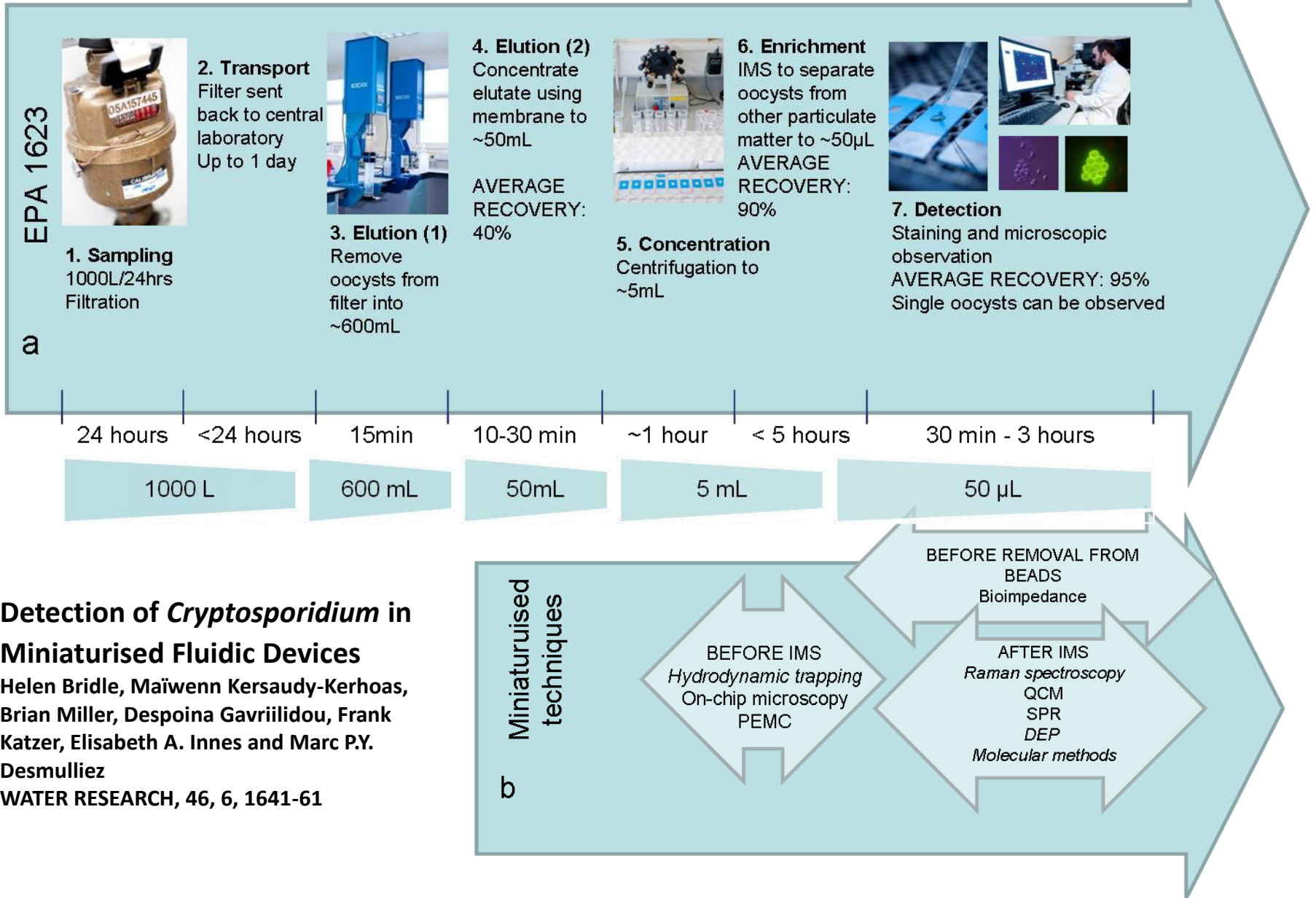
What?



	Pathogen	Equivalent diameter nm	
VIRUS	<i>Enteroviruses</i>	20-30	
	<i>Hepatitis E</i>	27-34	
	<i>Rotaviruses</i>	60-80	
	<i>Adenoviruses</i>	70-100	
	<i>Campylobacter spp</i>	310-1800	
BACTERIA	<i>Shigella spp</i>	510-2100	
	<i>E. coli</i>	720-1000	
	<i>Vibrio Cholera</i>	810-1400	
	<i>Samonella spp</i>	1100-2600	
PROTO.	<i>Giardia Lamblia</i>	2800-4600	
	<i>Cryptosporidium</i>	3600-7400	



DETECTION SYSTEMS



Detection of *Cryptosporidium* in Miniaturised Fluidic Devices

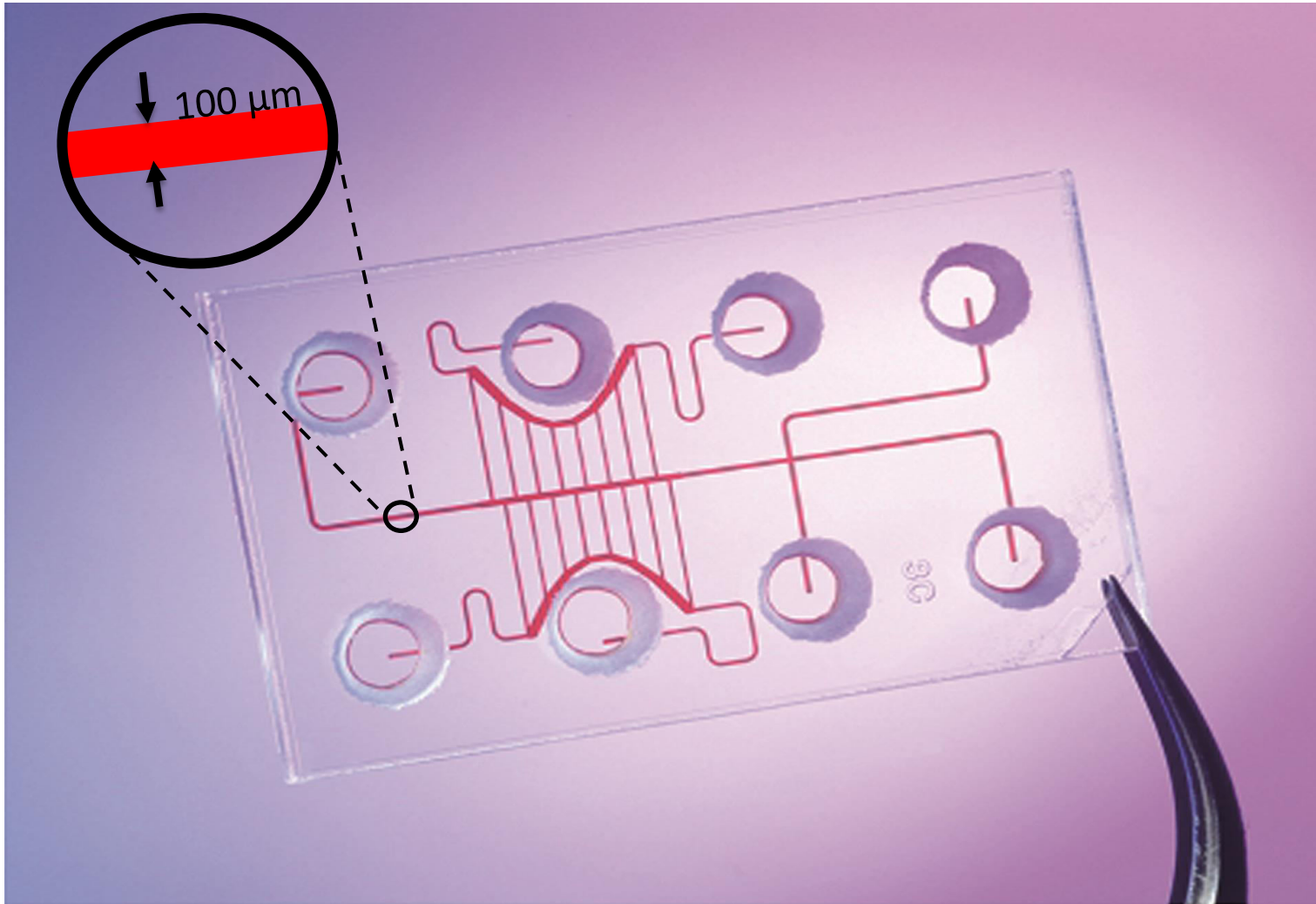
Helen Bridle, Maiwenn Kersaudy-Kerhoas, Brian Miller, Despoina Gavriilidou, Frank Katzer, Elisabeth A. Innes and Marc P.Y. Desmulliez

WATER RESEARCH, 46, 6, 1641-61

Challenges and Solutions

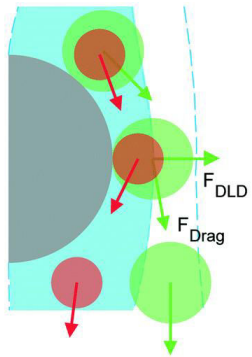
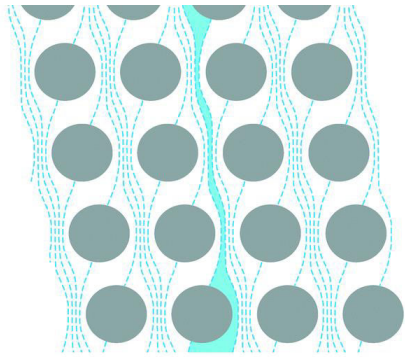
Challenges in Monitoring	Our Research
Recovery rates – sample processing challenge	Filtration systems – automated systems
	Filters – new materials
	Microfluidic approaches
Information obtained from detection	Active microfluidic separations
	Cantilever sensors
	Molecular methods
	Raman spectroscopy
Rapid online testing	Microfluidic early warning system

Microfluidics – Sample Processing and Detection

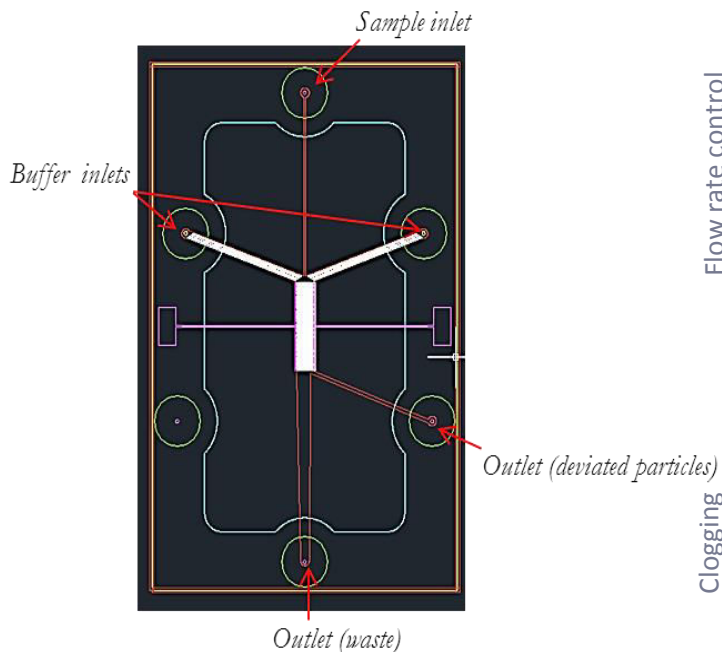


Bridle, H., Miller, B. and Desmulliez, M.P.Y. (2014) Application of microfluidics in waterborne pathogen monitoring: A review. *Water Research*, 55, 265-271.

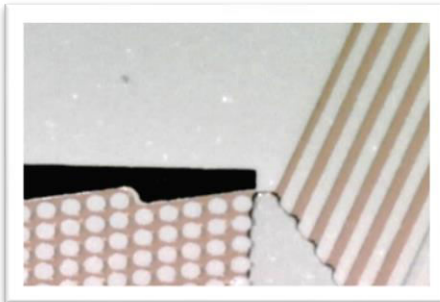
Passive Microfluidic Sample Processing



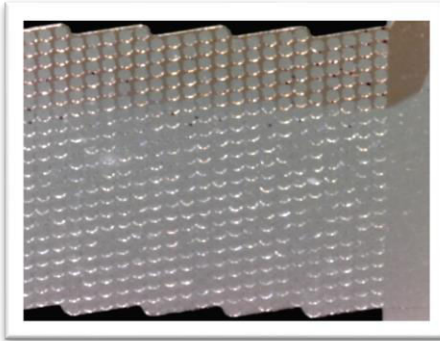
Beech et al, Lab Chip, 2009,
DOI: 10.1039/B823275J



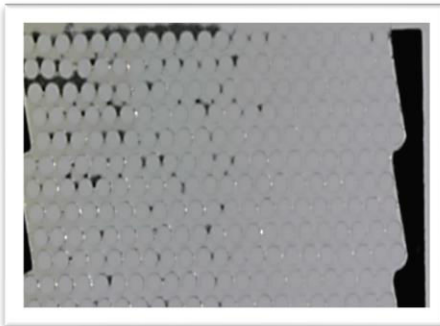
Bubbles



Flow rate control



Clogging



Jimenez and Bridle (OUTREACH)
Lab Chip, 2014,
DOI: 10.1039/C4LC00944D

McGrath, Jimenez and Bridle (REVIEW)
Lab Chip, 2014
DOI: 10.1039/C4LC00939H

John McGrath, Melanie Jimenez
Collaboration with Andras Laki,
PPKE, Budapest

Passive Microfluidic Sample Processing

ALTERNATIVE SYSTEM – several scales of performance

Protozoan Concentration Device

5mL input volume

Up to ~1mL/min

Run with spiked DI water, tap water, concentrate from tap water filtration and surface water filtration

Concentration factor equals number of outlets (2 outlets gives 2.5mL and 4 outlets gives 1.25mL from 5mL)

Increase concentration by recirculation and device stacking: 50mL to 1mL in ~10mins with a stack of 10 4 outlet devices recirculating 3 times

Filtration Type Systems

Stack of 20 larger systems operated at 1L/min

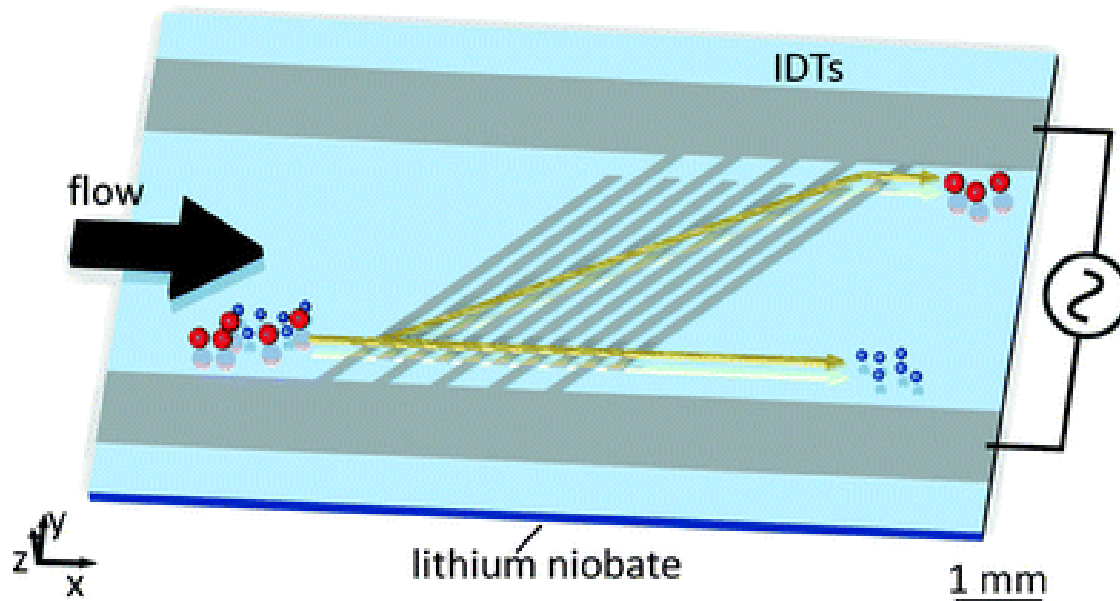
Filtered out larger particles (above ~75 μ m)

Single devices tested with tap and surface water

Brian Miller

PATENT about to be filed

ACTIVE FORCES FOR SEPARATIONS



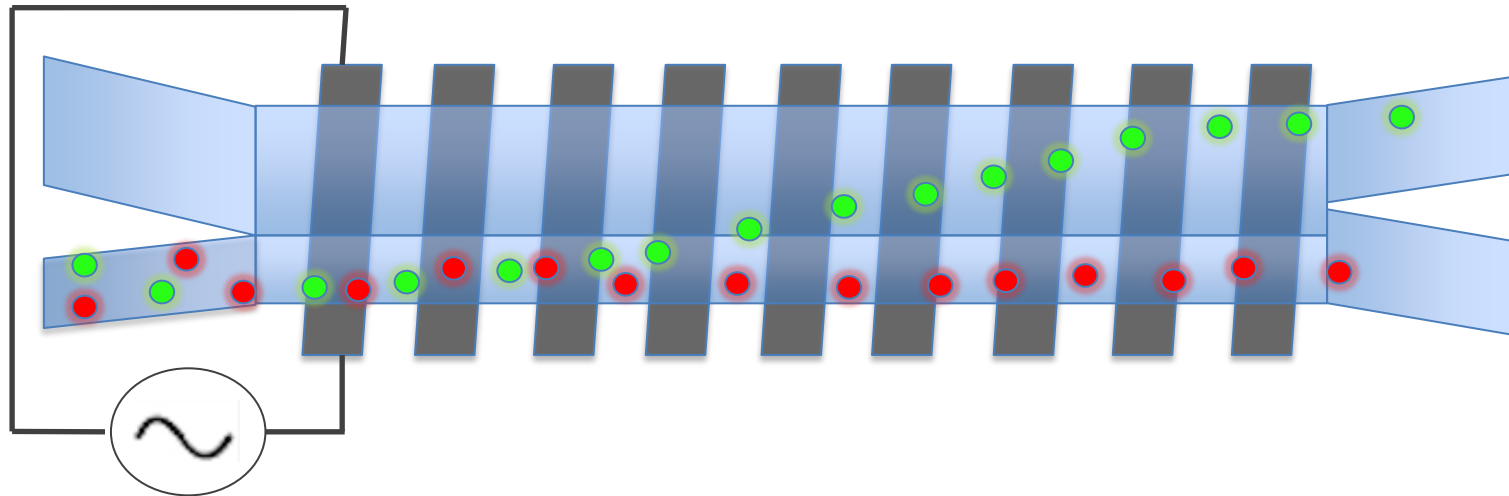
- Generate an acoustic force in the channel
- Size based separations

Collins, et al, Lab Chip, 2014, DOI: 10.1039/C3LC51367J

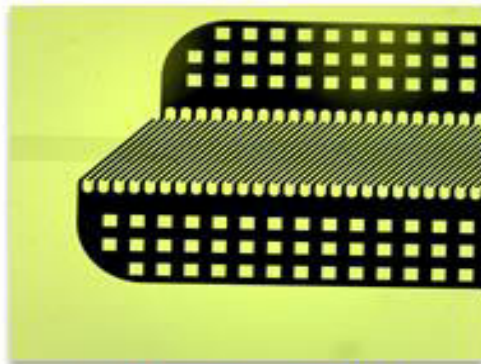
- Trialled the system with protozoa but encountered problems of sticking to the electrodes so a redesign has been produced and is awaiting testing

John McGrath
Dr Adrian Neild, Monash University

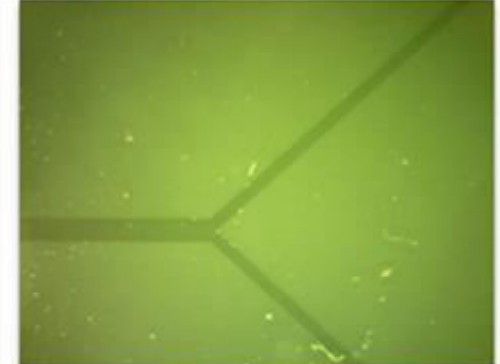
ACTIVE FORCES FOR SEPARATIONS



A. Wide buffer inlet enables focusing of sample inlet stream

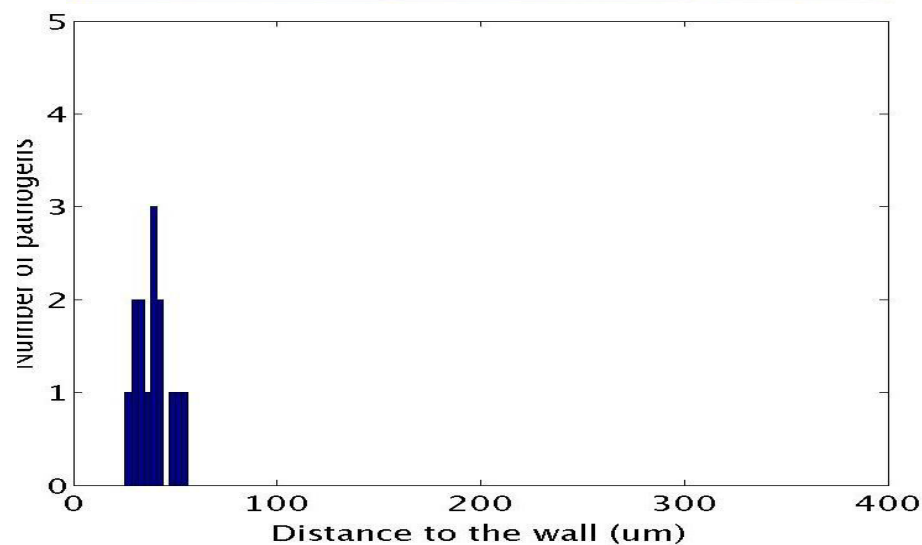
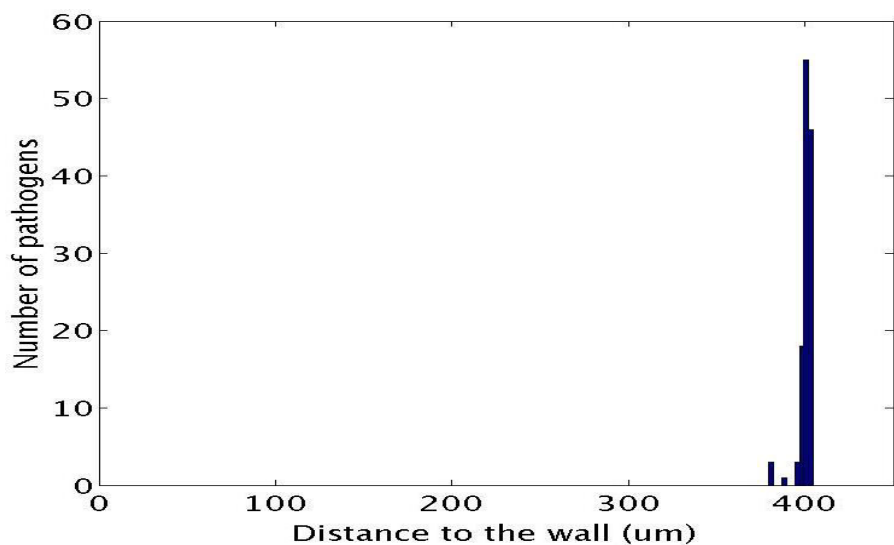
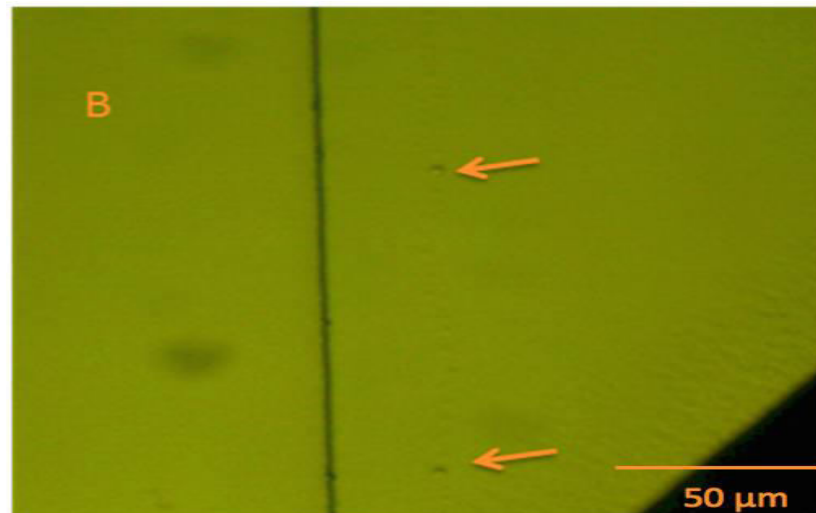


B. Electrodes at 45° to allow separation in main channel



C. Main channel separates into two equal divisions

ACTIVE FORCES FOR SEPARATIONS



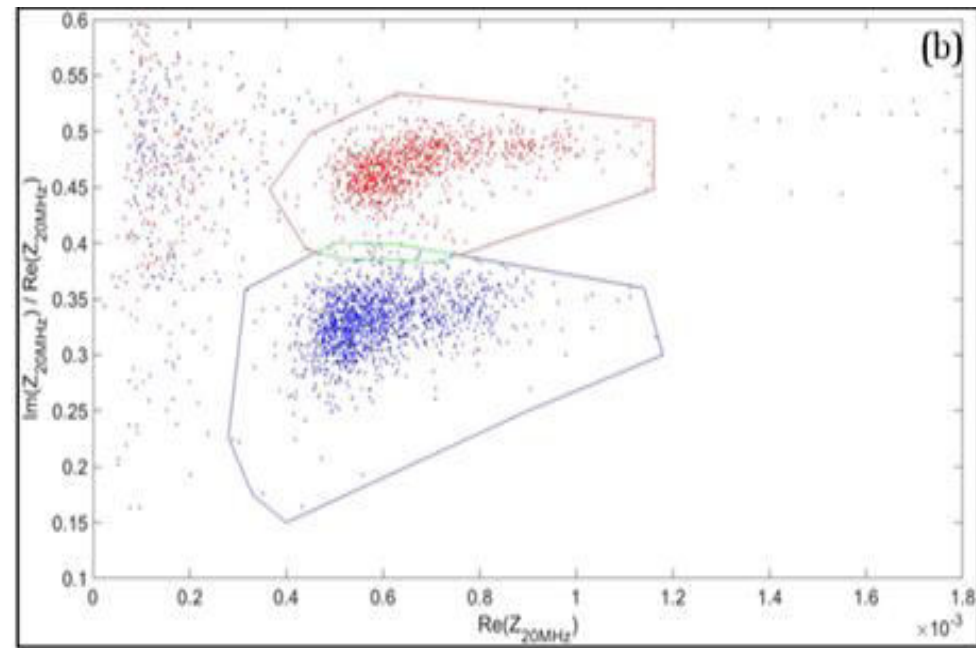
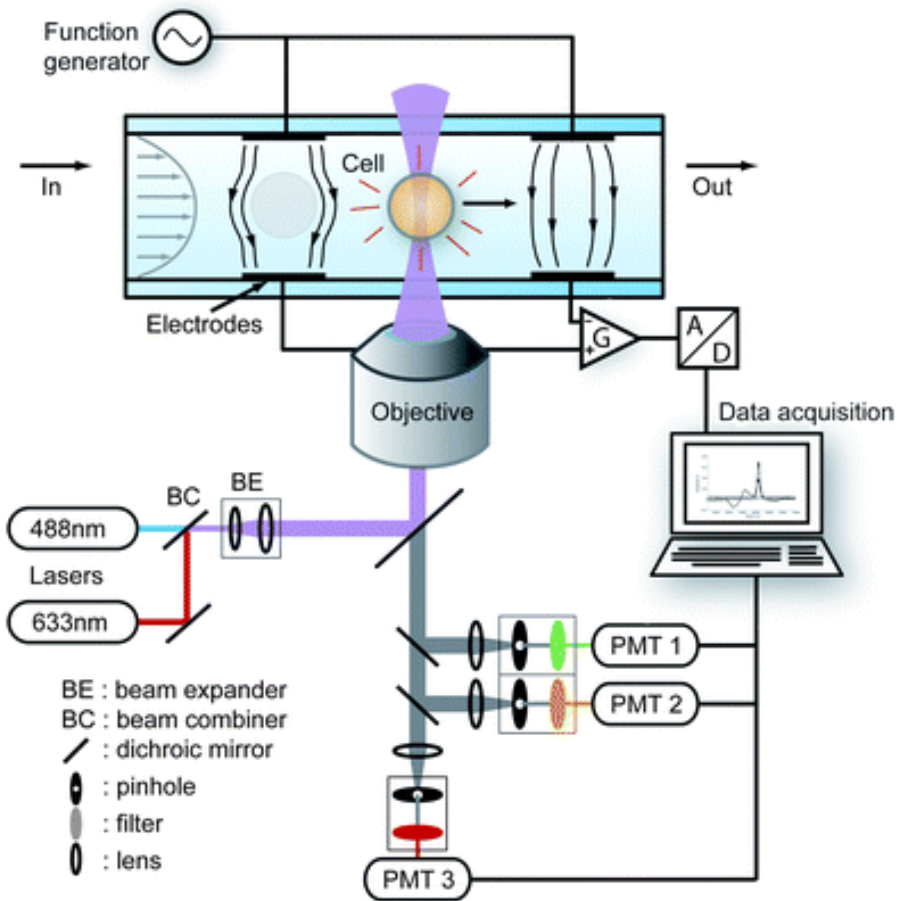
Displacement at 14 - 15 MHz, 12V

Sample and buffer introduced at $1 \mu\text{l min}^{-1}$ and $10 \mu\text{l min}^{-1}$, respectively

IMPEDANCE CYTOMETRY

Holmes et al, Lab Chip 2009

DOI: 10.1039/B910053A



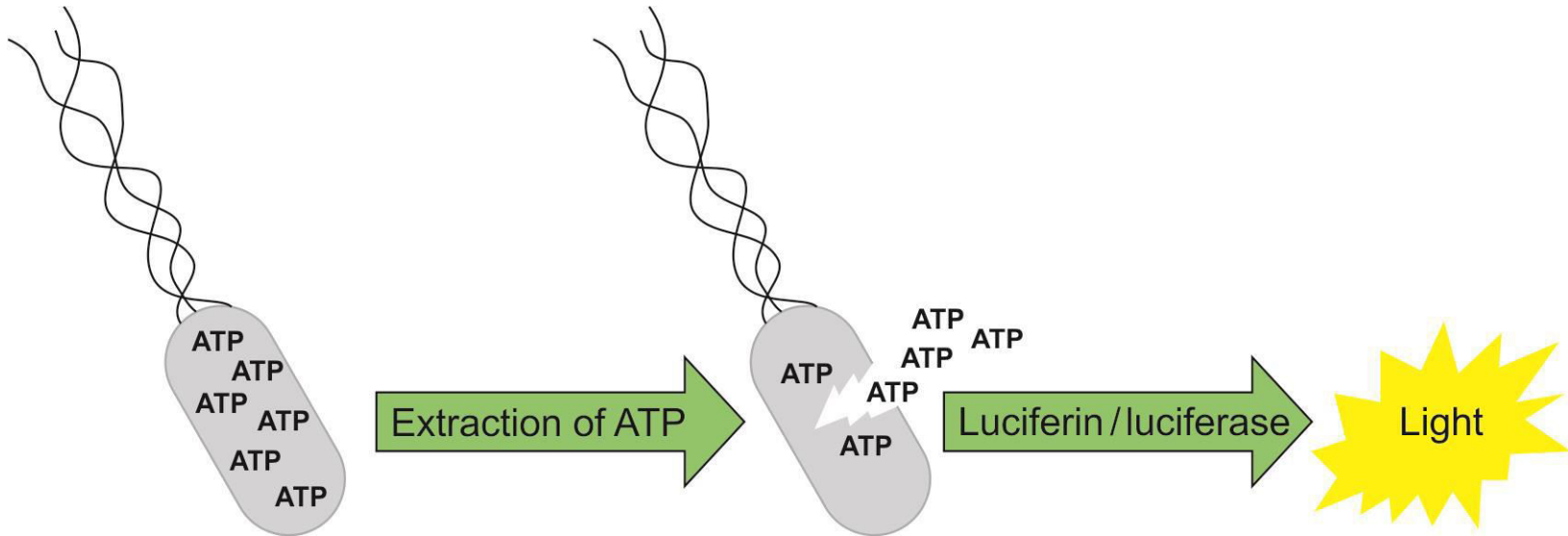
Discriminate viable and non-viable *C. parvum* (99% accuracy)

Discriminate between *C. parvum*, *C. muris* and *Giardia*

John McGrath

Professor Hywel Morgan, University of Southampton

Early Warning System



- Established method of detection which is applied for monitoring within the Netherlands
- Has been tested with drinking water spiked with wastewater samples achieving sensitivity comparable to total direct counts
- Sensitivity? 10⁻¹⁴ M ATP, 0.2 pg/mL

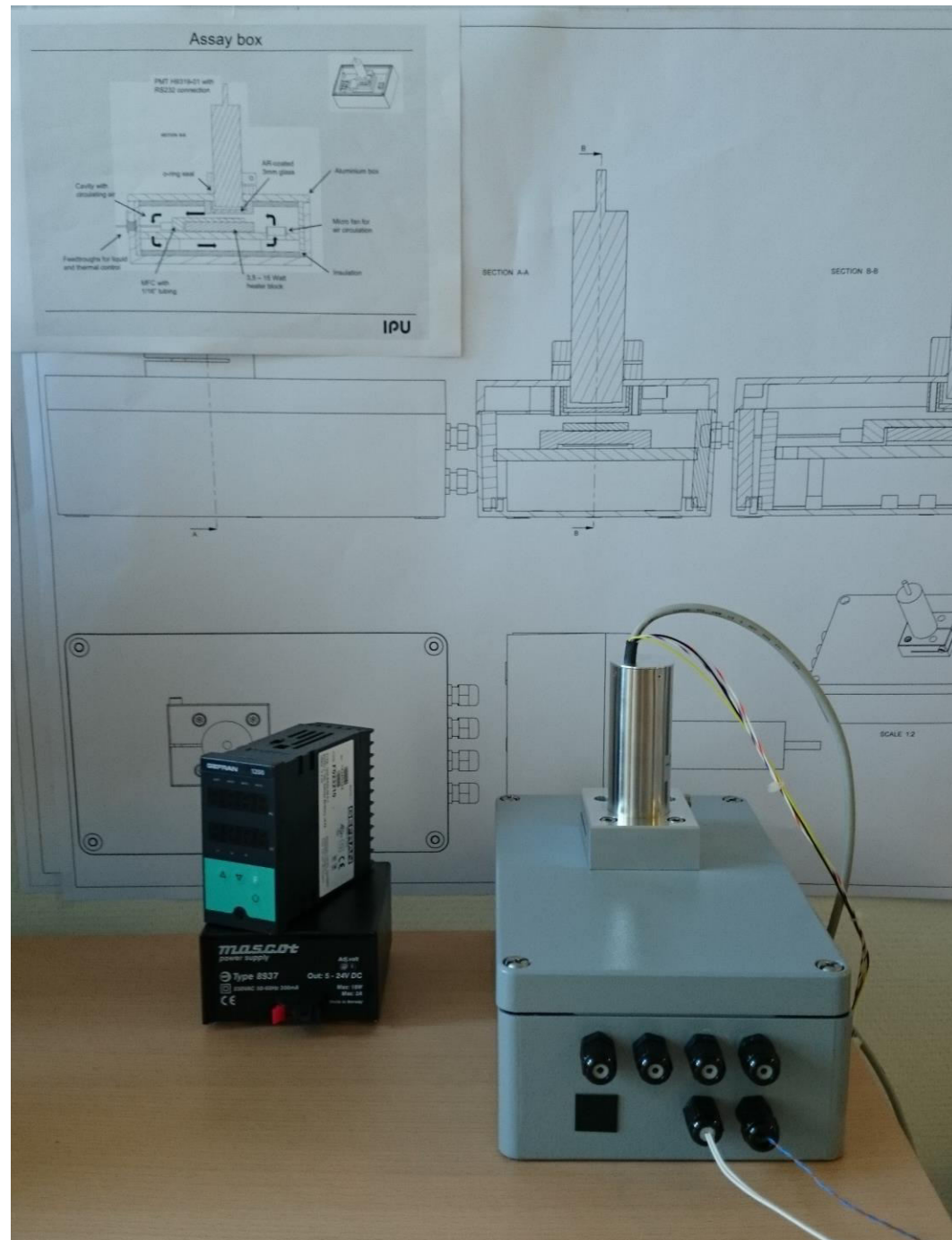
Vang Ó.K., C.B. Corfitzen, C. Smith & H.-J. Albrechtsen, 2014: Evaluation of ATP measurements to detect microbial ingress in drinking water by waste water and surface water **Water Research**, 64, 309-320.

Early Warning System



From lab equipment to
online testing unit

Abdelfateh Kerrouche
In collaboration with
Epigem and Claus
Barholm-Hanson (DTU
Environment)

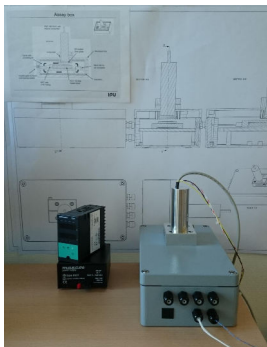
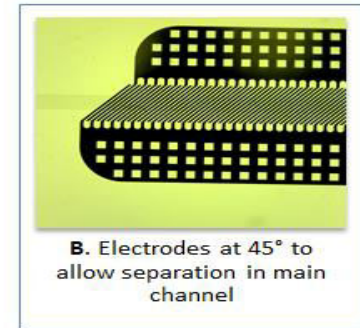
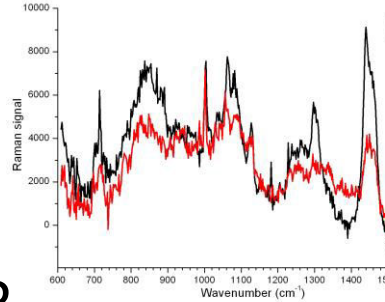
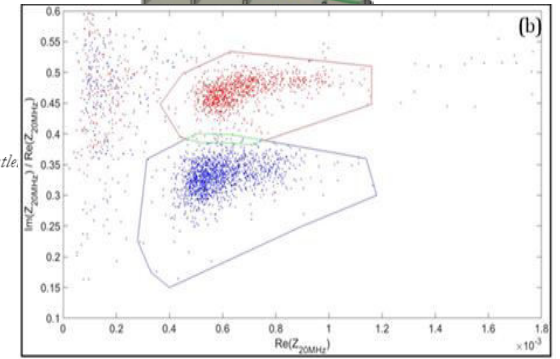
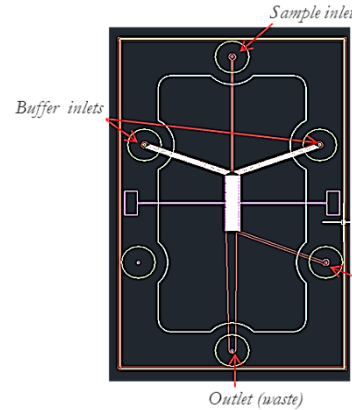
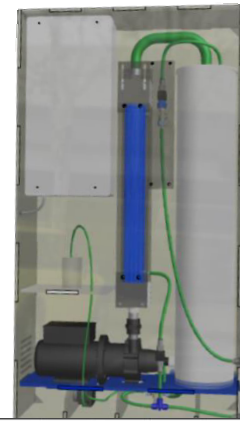
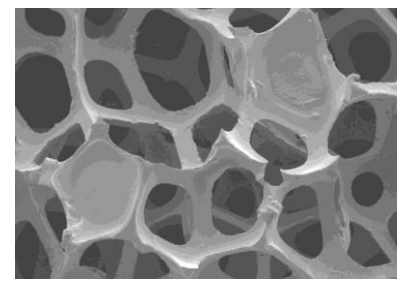
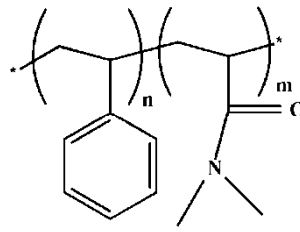


SUMMARY

SAMPLE PROCESSING –
automated filtration units, novel
filter materials and **microfluidic
solutions**

INFORMATION RICH DETECTION
– **microfluidics**, molecular
methods and Raman
spectroscopy

EARLY WARNING –
microfluidic online ATP



=> Range of easy to use high performance
monitoring technologies for safe drinking water

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Andras Laki, PPKE, Budapest

Professor Hywel Morgan, University of Southampton
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